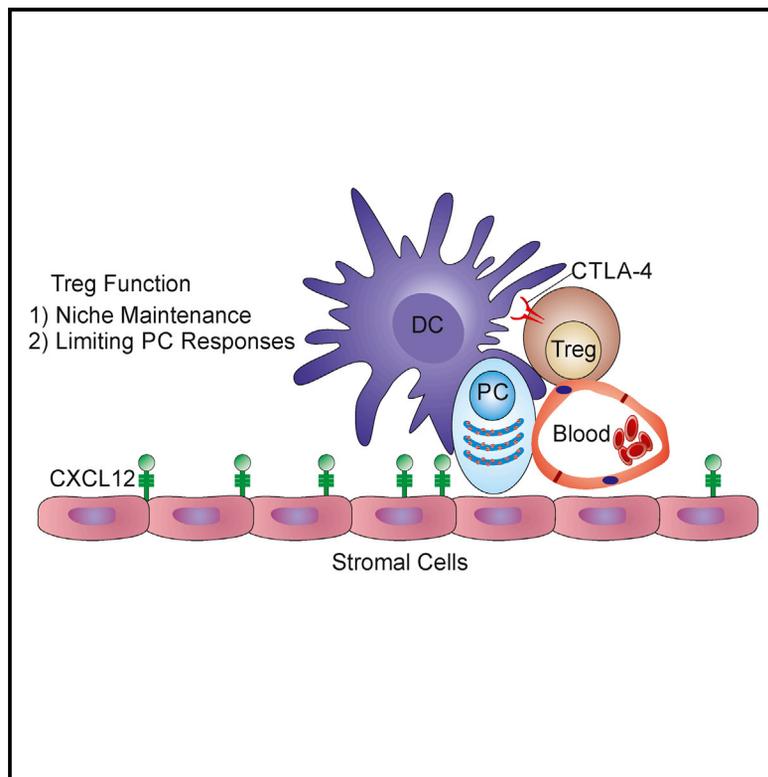


## T Regulatory Cells Support Plasma Cell Populations in the Bone Marrow

### Graphical Abstract



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### In Brief

Glatman Zaretsky et al. find that long-lived PCs in the bone marrow share a niche with Treg cells and dendritic cells. Their data suggest a role for Treg cells in the maintenance and regulation of PC populations. Understanding PC-Treg cell interactions could provide strategies to treat immune-mediated diseases with PC involvement.

### Highlights

- Systemic infection induces a loss of plasma cells and Treg cells in the bone marrow
- PCs and Treg cells share a niche with CD11c<sup>+</sup> cells
- Treg cells contribute to the regulation of PCs through CTLA-4
- Treg cells promote the maintenance of long-lived PCs in the BM

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# T Regulatory Cells Support Plasma Cell Populations in the Bone Marrow

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## SUMMARY

Long-lived plasma cells (PCs) in the bone marrow (BM) are a critical source of antibodies after infection or vaccination, but questions remain about the factors that control PCs. We found that systemic infection alters the BM, greatly reducing PCs and regulatory T (Treg) cells, a population that contributes to immune privilege in the BM. The use of intravital imaging revealed that BM Treg cells display a distinct behavior characterized by sustained co-localization with PCs and CD11c-YFP<sup>+</sup> cells. Gene expression profiling indicated that BM Treg cells express high levels of Treg effector molecules, and CTLA-4 deletion in these cells resulted in elevated PCs. Furthermore, preservation of Treg cells during systemic infection prevents PC loss, while Treg cell depletion in uninfected mice reduced PC populations. These studies suggest a role for Treg cells in PC biology and provide a potential target for the modulation of PCs during vaccine-induced humoral responses or autoimmunity.

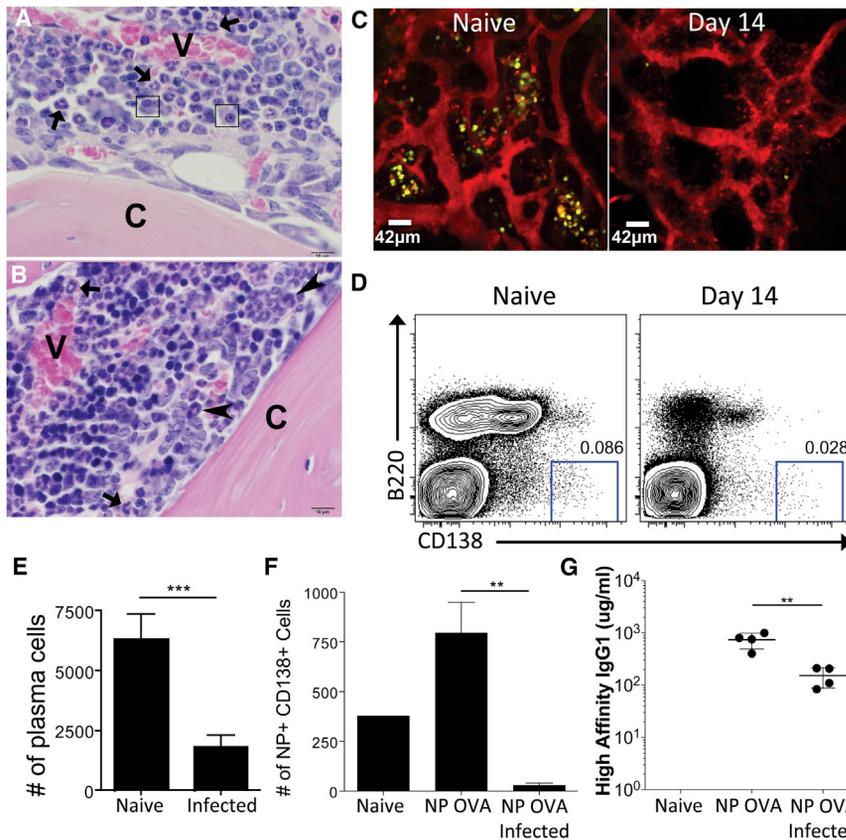
## INTRODUCTION

A variety of immune cell precursors reside and develop in the bone marrow (BM), a site that is also home to several populations of mature lymphocytes. There are multiple mechanisms to allow pluripotent or long-lived cells, including hematopoietic and cancer stem cells, plasma cells (PCs), and memory T cells, to persist in the BM (Fujisaki et al., 2011; Kawano et al., 2015). However, the spatial relationship and interactions between these disparate cellular populations are still being defined. For example, BM stromal cells provide growth and survival factors necessary for PC and hematopoietic stem cell (HSC) maintenance, but the

relationship between these niches is unclear (Sugiyama et al., 2006; Zehentmeier et al., 2014). Moreover, in the BM, regulatory T cells (Treg) are enriched and may contribute to the maintenance of the BM as an immune privileged site, necessary for HSC survival (Fujisaki et al., 2011). However, the behavior of Treg cells in the BM and their interactions with other immune populations have not been visualized and it remains unclear whether their activity is relevant to other hematopoietic cell populations in the BM.

Long-lived PCs present in the BM constitutively produce high levels of antibodies that result in life long serum antibody titers against previously encountered pathogens or vaccines (Manz et al., 1997; Slifka et al., 1998). Consequently, there is interest in understanding the mechanisms that maintain these cells (Chu and Berek, 2013). It is known that stromal cells provide survival signals to PCs through the production of CXCL13, BlyS, APRIL, and IL-6 (Roth et al., 2014). Furthermore, eosinophils, basophils, and megakaryocytes are implicated in the maintenance of PCs in the BM (Chu et al., 2011; Rodriguez Gomez et al., 2010; Winter et al., 2010), and there is evidence that perivascular clusters of DCs in the BM provide critical signals for B cells (Rozanski et al., 2011; Sapozhnikov et al., 2008). Although these factors promote PC survival, they are not sufficient, and the cellular composition of this niche and requirements for PC maintenance are major questions (Chu and Berek, 2013). However, there is a paucity of intravital imaging studies to describe the behavior of PCs and their interactions with other cell populations. Thus, there remains a need to better define the composition of this niche to understand how PCs are maintained and whether there are regulatory networks that limit PC responses.

Many studies have demonstrated that systemic infection or inflammation results in marked changes in BM populations (Glatman Zaretsky et al., 2012; MacNamara et al., 2009; Ueda et al., 2005). Here, challenge with *Toxoplasma gondii*, a clinically relevant parasite for which the mouse is a natural host (Montoya and Liesenfeld, 2004), demonstrated that infection-induced



**Figure 1. Acute *Toxoplasma gondii* Infection Results in a Loss of PCs in the BM**

(A) Naive mouse. The marrow cavity contains vascular sinuses (V) surrounded by mature neutrophils (arrowheads) admixed with predominantly myelopoietic precursors and few mature PCs (arrows). Bone cortex (C).

(B) Day 14 *T. gondii* infected mouse. Medullary vascular sinuses (V) are surrounded by increased numbers of hematopoietic progenitors characterized by hyperchromatic nuclei. Few mature neutrophils (arrowheads) and immature band neutrophils (arrows) are observed. Mature PCs are not identified. Bone cortex (C).

(C) Naive or infected BLIMP1-YFP reporter mice were imaged using intravital 2-photon microscopy of the skull BM. The BLIMP1-YFP-expressing cells are yellow and quantum dots were injected intravenously to label the vasculature red. At least three mice were imaged for each time point.

(D and E) BM from naive or infected WT mice was evaluated by flow cytometry (using a “dump” gate to eliminate CD3<sup>+</sup>, F4/80<sup>+</sup>, and/or Gr1<sup>+</sup> potential contaminating cells) for the presence of PCs.

(F and G) WT mice were immunized with NP-OVA. Six weeks later, these mice were challenged with *T. gondii* and evaluated for NP-specific PCs (F) and NP-specific antibody titers (G).

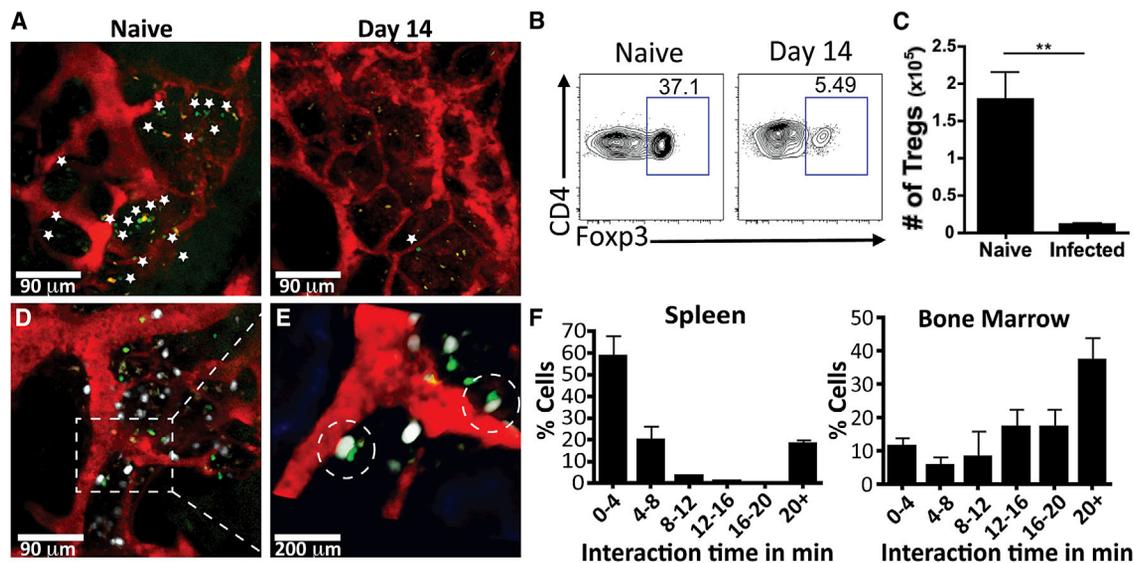
changes in the BM lead to a transient loss of PCs and Tregs. Multiphoton imaging revealed that at homeostasis, PCs interact with a CD11c-YFP<sup>+</sup> population and Treg cells in the BM. Further studies indicate that Treg cells have a complex interaction with BM PCs, including a CTLA-4-dependent mechanism of PC regulation, while supporting PC populations. Together, these studies suggest an unanticipated role for Treg cells in the maintenance and operation of the PC niche within the BM.

## RESULTS

### Plasma Cells in the BM Are Disrupted by Systemic Infection

Changes in the BM, including increased hematopoiesis and reduced lymphopoiesis, are a hallmark of the immune response to numerous pathogens (Glatman Zaretsky et al., 2014; Manz and Boettcher, 2014). Indeed, following infection with the systemic pathogen *T. gondii*, histological analysis of the BM revealed extensive changes in the cellular composition of this compartment, including increased numbers of hematopoietic progenitors (Figures 1A and 1B). Unexpectedly, resident PC populations, identified based on their oval shape, purple cytoplasm, and characteristic eccentric nucleus with clock-face chromatin and perinuclear clearing, were diminished (large arrows) (Figures 1A and 1B). Long-lived PC populations are an independent pool of terminally differentiated B cells maintained through slow homeostatic proliferation with a half-life of >120 days (Slifka et al., 1998). PCs express high levels of

the transcription factor BLIMP-1 (Minnich et al., 2016; Tellier et al., 2016; Turner et al., 1994) and in uninfected BLIMP-YFP or BLIMP-GFP mice, 70%–85% of the CD138<sup>+</sup> PCs were positive for the reporter, which indicated sufficient fidelity to visualize PC populations (Wolf et al., 2011). Therefore, intravital imaging of the BM in the skull of BLIMP1-YFP reporter mice was used to visualize the location and motility of PC populations in uninfected and infected mice (Figure 1C). In naive mice, PCs are localized in clusters adjacent to blood vessels in the endosteal region of the bone and were stationary over the imaging periods used in these studies (Movie S1). However, by 14 days post infection, the PC population was markedly reduced (Figure 1C), and flow cytometry confirmed the infection-induced loss of PCs (Figures 1D and 1E). This analysis also revealed a decrease of >95% in the total number of pro, pre, immature, and mature recirculating B cells (Figures S1A–S1C). Although short-term blockade of B cell lymphopoiesis has minimal effects on mature B cell compartments (Hao and Rajewsky, 2001), it was possible that these alterations might impact the development of PC populations. To ascertain whether infection reduced an established PC population, mice were immunized with 4-hydroxy-3-nitrophenylacetyl (NP)-ovalbumin. This antigen results in a long-lived NP-specific PC population in the BM, detectable by intracellular staining with NP conjugated to allophycocyanin (Figure 1F). During acute *T. gondii* infection, the number of NP-specific B cells in the spleen was unchanged (data not shown), but there was a decrease in the number of NP<sup>+</sup> PCs in the BM, accompanied by a significant drop in serum NP-specific IgG1 (Figures 1F and 1G). However, by the chronic phase of infection, the NP<sup>+</sup>



**Figure 2. Treg Cells and PCs Interact in the BM**

(A) Naive or infected Foxp3-GFP reporter mice were imaged using intravital 2-photon microscopy of the skull BM. The Foxp3-GFP-expressing cells are green (demarcated by the \*) and quantum dots were injected intravenously to label the vasculature red. At least three mice were imaged for each time point.

(B and C) BM from naive or infected WT mice was evaluated by flow cytometry for the presence of Treg cells.

(D–F) BLIMP1-YFP/Foxp3-GFP dual reporter mice were imaged using intravital 2-photon microscopy of the skull BM. (D and E) BLIMP1-YFP (white), Foxp3-GFP (green), and vasculature (red) expression in BLIMP1-YFP/Foxp3-GFP dual reporter mice reveals short and long term interactions, highlighted in the close up (E). Three mice were imaged for these experiments, with 53 Foxp3-GFP cells recorded in the BM and 104 imaged in the spleen. (F) Quantification of the duration of PC-Treg cell interactions in the spleen and BM.

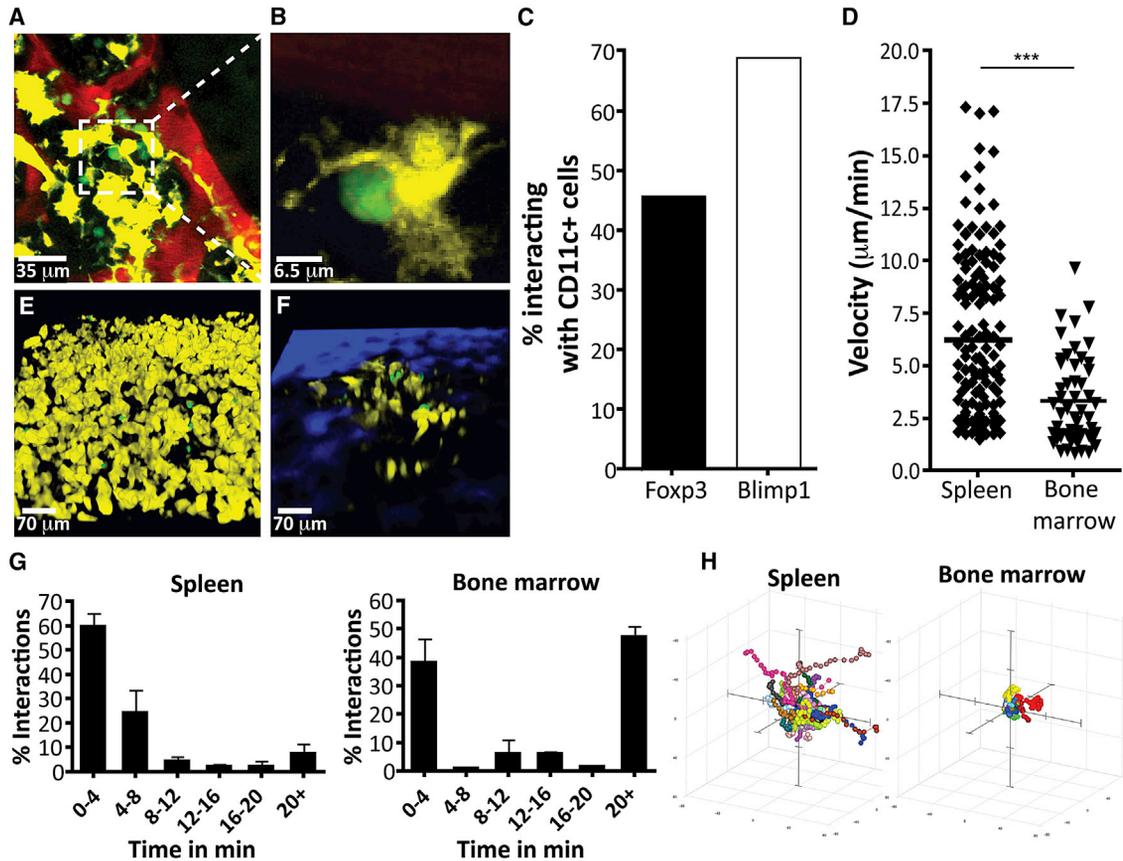
population was restored to its original levels (data not shown) indicating that infection results in the transient loss of a pre-established PC compartment.

### Plasma Cells Interact with T Regulatory Cells and CD11c<sup>+</sup> Cells in the BM

The infection-induced loss of PCs in the BM raises questions about which of the regulatory or cellular elements that support the PC niche are disrupted. Because Foxp3-GFP<sup>+</sup> cells are preferentially localized in the endosteal region of naive BM and contribute to the status of the BM as an immune privileged site (Fujisaki et al., 2011), we examined the impact of infection on this population. Intravital imaging of uninfected mice showed Foxp3-GFP<sup>+</sup> Treg cells throughout the BM, predominantly in extravascular sites, but by day 14 post-infection, these cells were largely absent (Figure 2A). Flow cytometric analysis confirmed that infection resulted in an ~80% decrease in Treg cells in the BM (Figures 2B and 2C) that mirrored the changes in Treg cell numbers at peripheral sites during acute toxoplasmosis (Benson et al., 2012; Oldenhove et al., 2009). The generation of mice that express both BLIMP1-YFP and Foxp3-GFP allowed for simultaneous imaging of PC and Treg cell populations in the BM. While Treg cells exhibited a greater motility than the PCs, these cells have a range of mobility with a large proportion of the Treg cell population displaying a more sessile phenotype (Movie S2). Indeed, the majority of the Foxp3-GFP<sup>+</sup> cells were found closely associated with BLIMP1-YFP<sup>+</sup> PCs (Figures 2D–2F), as ~10% of PCs interact with Treg cells for 4 min or less, while more than 60% of interactions between Foxp3-GFP<sup>+</sup>

and BLIMP1-YFP<sup>+</sup> cells were >10 min, with 30% exceeding the 20-min imaging window (Figure 2F; Movies S3 and S4). In contrast, Foxp3-GFP<sup>+</sup> cells in the spleen exhibited a higher velocity and shorter interaction times with BLIMP1-YFP<sup>+</sup> cells (Figure 2F; Movie S5). It should be noted that the transfer of polyclonally activated CD8<sup>+</sup> T cells into Foxp3-GFP reporter mice resulted in a small population of CD8<sup>+</sup> T cells in the BM that could be imaged simultaneously with Treg cells. These effector T cells exhibited a higher velocity than that of the resident Treg cell population (Figure S2A; Movie S6). This internal control indicates that the sessile Treg cell phenotype in the BM was not an artifact associated with sample preparation. Further analysis of the cellular localization of PCs and Treg cells using fixed long bones showed that they were present in areas rich in HSCs (CD150<sup>+</sup>), and PCs were intimately associated with the network of stromal cells that are a source of CXCL12 in the BM (Figures S2B and S2C; Movie S7). Thus, Treg cells in the BM display a behavior distinct from that observed in the spleen and many are spatially associated with PCs within local niches in the BM.

Previous studies have described a population of mature recirculating B cells in the BM that are closely associated with DCs (Sapozhnikov et al., 2008), but it is unclear whether DCs are a component of the PC niche. Therefore, mice that express CD11c-YFP/BLIMP1-GFP or CD11c-YFP/Foxp3-GFP were generated to determine whether DCs are localized near to or interact with PCs or Treg cells in the BM. In the cavarial BM of naive CD11c-YFP/BLIMP1-GFP mice, there were numerous large stationary CD11c-YFP<sup>+</sup> populations that displayed irregularly shaped dendrites (Figures 3A and 3B),



**Figure 3. CD11c-YFP<sup>+</sup> Cells Interact with PCs and Treg Cells in the BM**

(A–C) Intravital imaging of skull BM in BLIMP-GFP/CD11c-YFP dual reporter mice with quantum dot-labeled vasculature (red), with interacting cells highlighted in close up (B). Four mice were imaged, for a total of 83 BLIMP-GFP cells counted for quantification. (C) Quantification of the percentage of cells observed in interactions between CD11c-YFP<sup>+</sup> cells and BLIMP1-GFP<sup>+</sup> or Foxp3-GFP<sup>+</sup> cells in respective dual reporter mice.

(D–H) Imaging and quantification of Treg cell velocity (D) and CD11c<sup>+</sup> cell-Treg cell location (E and F) in the spleen and BM of Foxp3-GFP (53 BM Foxp3-GFP cells and 150 splenic Foxp3-GFP cells) (D) or CD11c-YFP/Foxp3-GFP dual (E–G) reporter mice, as well as interaction times between these cells (68 BM Foxp3-GFP cells and 94 splenic Foxp3-GFP cells) (G) and Treg cell movement shown by tracks (H).

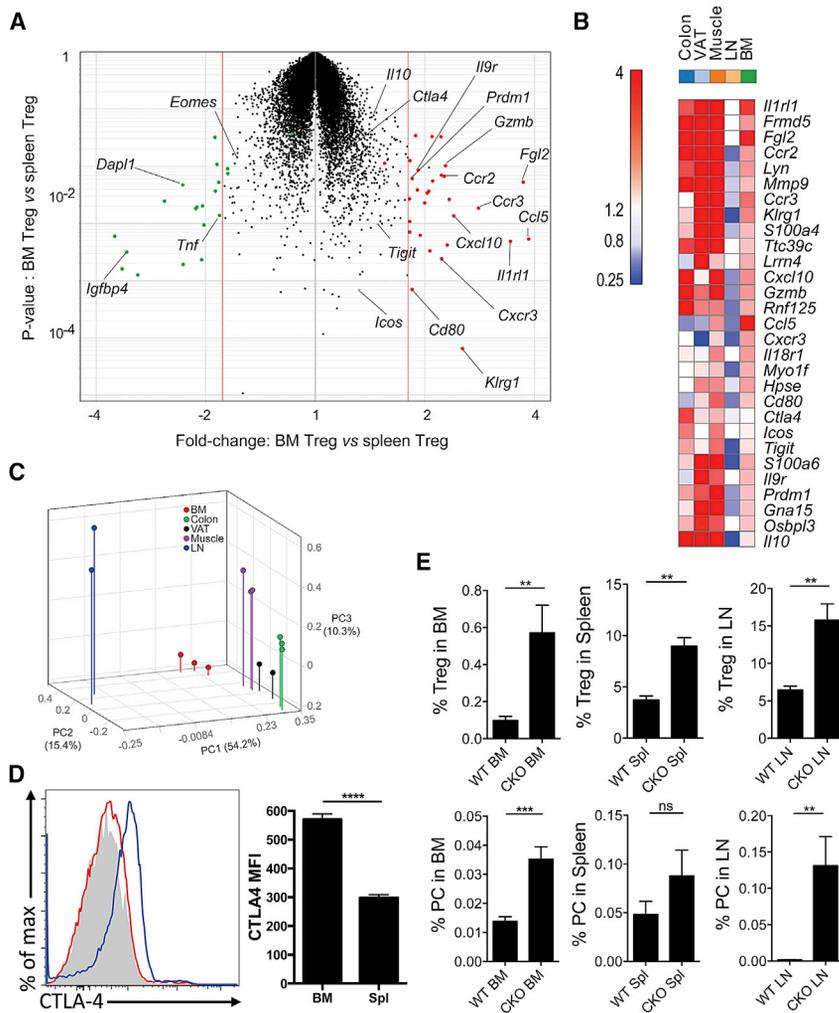
and ~70% of PCs interacted with a population of CD11c-YFP<sup>+</sup> cells, while 40%–50% of Treg cells interacted with CD11c-YFP<sup>+</sup> cells (Figure 3C; Movie S8). The use of mixed BM chimeras demonstrated that the CD11c-YFP<sup>+</sup> cells present in the BM are of hematopoietic origin, as wild-type (WT) recipients that were reconstituted with BM from CD11c-YFP mice contained an extensive network of YFP<sup>+</sup> cells in the BM, while YFP<sup>+</sup> cells were rare in irradiated reporter mice that were recipients of WT BM, associated with a 40-fold decrease in the YFP volume (Figure S3). These data support a model in which CD11c-YFP<sup>+</sup> cells, implicated in providing co-stimulation required for PC survival and antibody production (Rozanski et al., 2011), are a component of the PC niche and form close associations with PCs and Treg cells.

Few studies have used multi-photon imaging to visualize Treg cell behavior and interactions with DCs *in situ* and these were performed in LNs in the context of immunization and diabetes (Matheu et al., 2015; Tadokoro et al., 2006; Tang et al., 2006). Therefore, we decided to compare how Treg cells in the BM or spleen interact with resident DC populations at homeostasis.

Analysis of reporter mice revealed that splenic Treg cells were highly motile based on their velocity and displacement rate, and in the spleens of Foxp3-GFP/CD11c-YFP dual reporter mice, <10% of the Treg cells had sustained interactions with CD11c-YFP<sup>+</sup> cells (Figures 3D, 3E, 3G, and 3H; Movie S9). In contrast, Treg cells in the BM had reduced velocity, low displacement, and >50% of Treg cells had sustained interactions with CD11c-YFP<sup>+</sup> cells (Figures 3C, 3D, 3F–3H; Movie S10). Therefore, the Treg cell populations in the spleen and BM are characterized by fundamental differences in their behavior and those in the BM display unanticipated interactions with PCs, as well as with a CD11c-YFP<sup>+</sup> population.

### Treg Cells in the BM Exhibit an Effector Phenotype with Regulatory Effects on PCs

Treg cells in non-lymphoid tissues have unique transcriptional profiles and functions associated with individual tissues (Burzyn et al., 2013a, 2013b; Feuerer et al., 2010). However, despite the enrichment of Treg cells in the BM, these cells have not been transcriptionally characterized. Therefore, to determine whether



**Figure 4. BM Treg Cells Exhibit a More Effector-like Phenotype than Splenic Treg Cells with a Higher Expression of Suppressor Genes**

(A–C) Treg cells sorted from spleen and BM of naive Foxp3-GFP<sup>+</sup> mice were evaluated by microarray. Cells were assessed for differentially regulated genes (A), which were then compared to Treg cells from other sites by specific gene expression (B) or as total populations by principle component analysis (C).

(D) CTLA-4 expression was determined by flow cytometry on BM or splenic Treg cells, which was concatenated from five mice (left) and MFI was quantified (right).

(E) PC and Treg cell populations in the BM, Spl, and LN of WT or Foxp3<sup>Cre</sup>/CTLA4<sup>fl/fl</sup> (CKO) mice were assessed by flow cytometry.

focused on CTLA-4 as being highly expressed by BM Treg cells and having a prominent role in Treg suppressive activity (Wing et al., 2008). Analysis of CTLA-4 expression revealed that splenic Treg cells express negligible surface levels of CTLA-4, whereas BM Treg cells have elevated levels (Figure 4D). Next, we analyzed mice with a Treg cell-specific loss of CTLA-4 expression (Foxp3<sup>Cre</sup>/CTLA4<sup>fl/fl</sup>), which develop systemic inflammation (Wing et al., 2008). Despite the presence of an ongoing inflammatory response, there was no decrease in the numbers of Foxp3<sup>+</sup> T cells in the BM and these mice had a ~3-fold increase in the percentage of Treg cells (Figure 4E). However, in contrast to the loss of PCs

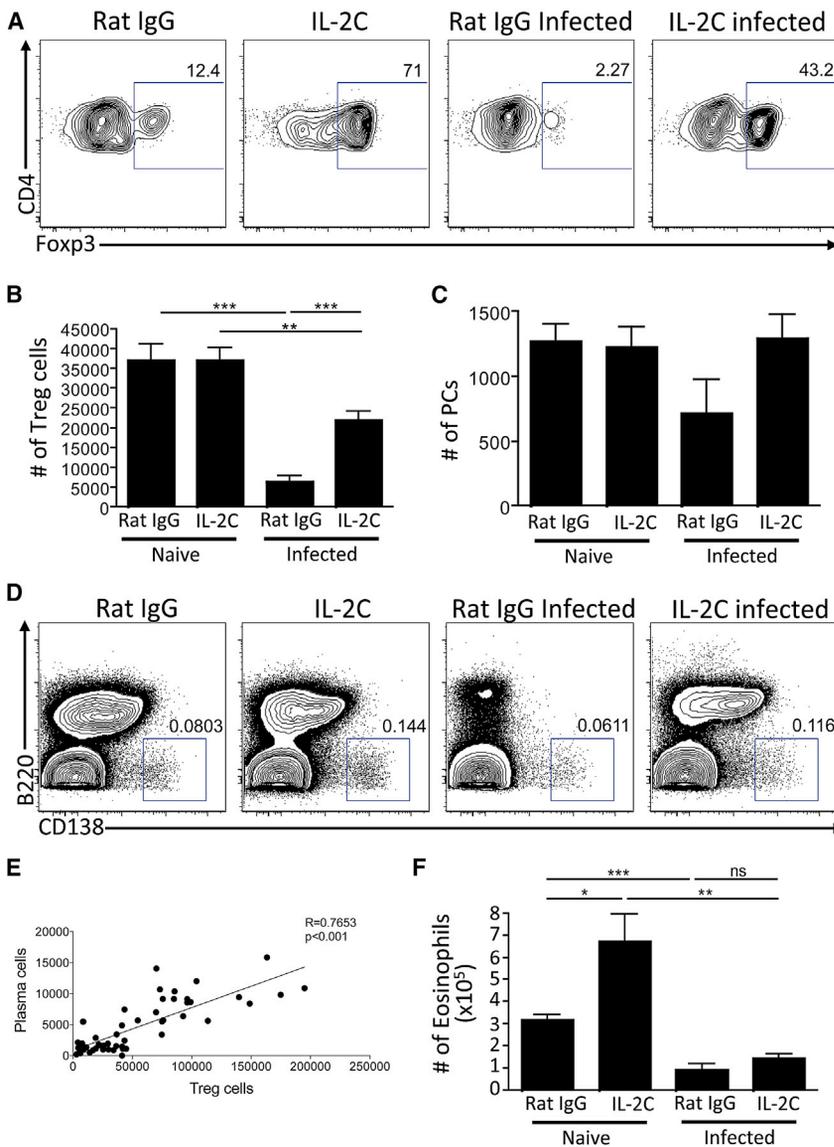
BM Treg cells have a transcriptional signature distinct from that of peripheral Treg cells, gene expression profiles of BM or splenic Treg cells were compared (Figure 4A; Table S1). Differentially expressed transcripts are dominated by chemokine/cytokine receptors (*Ccr2*, *Ccr3*, *Cxcr3*, *Il9r*, *Il1r1*) and effector molecules (*Ctla4*, *Il10*, *Fgl2*, *Gzmb*), suggestive of an activated population with increased Treg suppressive capabilities (Figures 4A and 4B). These transcripts are also overexpressed in other tissue-Treg cells relative to those isolated from lymphoid organs, with those from injured muscle being most similar to BM Treg cells (Figure 4B). Indeed, principal component analysis encompassing all differentially represented transcripts showed that BM Treg cells reside with tissue Treg cells, not lymphoid Treg cells (Figure 4C). These findings highlight the effector capacity of BM Treg cells, consistent with the idea that these cells have a role in the maintenance of an immune privileged niche.

Because PCs in the BM co-localize with Treg cells, it was relevant to determine whether the reduction in PCs during infection is a consequence of the loss of the BM Treg cell population or a secondary consequence of inflammation due to the loss of the suppressive niche. Based on the transcriptional profiling, we

observed when Treg cells are reduced during infection-induced inflammation, compromising Treg cell function in this system without decreasing Treg cell number resulted in a 3-fold increase in the number of PCs (Figure 4E). However, CTLA-4 expression on Treg cells negatively regulates PC development in the periphery (Sage et al., 2014; Wing et al., 2014), and PCs in the BM express high levels of CD28, an important regulator of PC function (Rozanski et al., 2011). Thus, although CTLA4-deficient Treg cells are unable to prevent fatal inflammation (Wing et al., 2008), these experiments suggest that Treg cell expression of CTLA-4 acts to limit the size of the PC pool in the BM.

### Treg Cells and PC Maintenance

Next, complementary approaches were used to understand whether Treg cells are directly involved in the maintenance of PCs. First, in gain-of-function experiments, we asked whether treatment with an interleukin-2 (IL-2)-anti-IL-2 antibody complex that promotes Treg cell survival (Boyman et al., 2006) could prevent the loss of BM PCs during infection. Treatment of naive mice with IL-2 complex resulted in elevated Treg cell numbers by percentage, but not absolute number in the BM (Figures 5A and 5B),



**Figure 5. BM Treg Cells Support PC Maintenance in the BM**

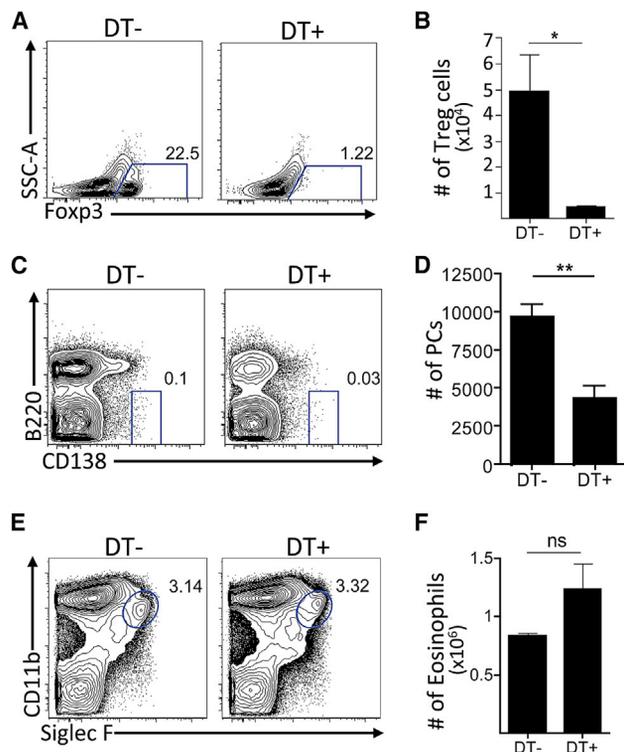
(A–F) WT mice were infected with *T. gondii* and treated with IL-2-anti-IL-2 complex every 3 days for 14 days. BM was evaluated for Treg cells (A and B), PCs (C and D), and representative flow plots are shown, with a concatenation of mice within each group for PC plots (D). The numbers of Treg cells and PCs were plotted to identify correlations between the cell numbers of these two populations (E). Eosinophils were evaluated and quantified by flow cytometry (F).

cells are transiently lost from the BM along with PCs during acute infection (Figure 5F). However, as previously reported (Van Gool et al., 2014), treatment of naive mice with the IL-2 complex resulted in an eosinophilia, but when infected mice were treated with IL-2 complex, the eosinophil population was not rescued, although the PC population was maintained (Figure 5F). These data suggest that the role for Treg cells in supporting PC populations is distinct from that of the eosinophils present in the BM.

Second, to assess the impact of Treg cell loss on PC populations in the absence of infection, Treg cells were depleted in naive Foxp3-DTR mice using diphtheria toxin (DT) and the PC compartment was assessed. In these mice, DT treatment resulted in a >90% decrease in the percentage and number of Foxp3<sup>+</sup> Treg cells in the BM (Figures 6A and 6B) and a >50% decrease in PCs (Figures 6C and 6D). Of note, when Treg cells were eliminated and PCs were reduced, the numbers of CD11c<sup>+</sup>MHCII<sup>hi</sup> cells remained constant, although these cells did express elevated levels of CD80 (data not shown). Thus,

and did not alter PC numbers (Figures 5C and 5D). IL-2 complex treatment of infected mice did result in a modest increase in parasite burden (data not shown), but mitigated the loss of the BM Treg cells and preserved the PCs in the BM (Figures 5A–5D). As PCs do not express the IL-2 receptor component CD25 (Jego et al., 2001) (data not shown), this rescue is unlikely due to direct effects of the IL-2-anti-IL-2 complex on PCs. Of note, because of the experimental variation we observed in the levels of PC reduction, the overall relationship between the frequency of Treg cells and PCs was assessed by comparing Treg cell numbers with the numbers of PCs in the BM from these experiments. When the number of Treg cells was plotted versus the number of PCs in the BM from these IL-2 treatment experiments, a highly significant correlation was observed between these populations (Figure 5E). Of note, it has been proposed that eosinophils promote PC survival (Chu et al., 2011) and these

despite recent findings that Treg cell depletion enhances PC formation in the periphery (Wing et al., 2014), the depletion of Treg cells resulted in a reduction in the BM PC population, similar to the phenomenon observed in infected mice. Although Treg cell depletion using the Foxp3-DTR system results in widespread inflammation (Christiaansen et al., 2014), the studies described in Figure 5 are performed in the context of systemic infection-induced inflammation. Thus, the ability of IL-2C treatment to preserve Treg cells and mitigate the PC loss suggests that inflammation alone would not result in the PC reduction observed in the Foxp3-DTR experiments. In these mice, Treg cell depletion did not result in significant changes in the eosinophil population, further emphasizing that eosinophils are not sufficient for PC maintenance (Figures 6E and 6F). These independent approaches implicate Treg cells as part of a network that maintains and regulates PC populations in the BM.



**Figure 6. A Loss of Treg Cells Correlates with a Reduced PC Population**

Naive Foxp3-DTR Mice Were Treated with DT on Days 0, 1, 4, and 6 and Evaluated on Day 7.

(A–F) Populations of Treg cells (A and B), PCs (C and D), and eosinophils (E and F) were evaluated and enumerated in the BM.

## DISCUSSION

There have been many advances in defining the events that lead to the generation of PCs and the idea that BM PCs require multiple survival factors for their maintenance is established, but the cellular interactions that promote PC survival are poorly characterized (Goodnow et al., 2010). Stromal cells are part of this niche (Tokoyoda et al., 2004; Zehentmeier et al., 2014), and diverse migratory cells in the BM influence PC survival (Tangye, 2011; Winter et al., 2012), and data presented here highlight the association of BM PCs and Treg cells with CD11c-YFP<sup>+</sup> cells. Together with evidence that Treg cells contribute to HSC survival (Fujisaki et al., 2011), our studies connecting Treg cells with maintenance of BM PCs suggest that Treg cell activities in the BM may affect the homeostasis of many cell types. Indeed, Treg cells appear to be in excess of the BM PCs, which may prevent small changes in Treg cell numbers from resulting in major changes in PC homeostasis. Alternatively, “excess” Treg cells may reflect their role in other niches (Fujisaki et al., 2011), and the principles that govern distinct micro-environments may offer insight into the mechanisms that preserve memory cells or which help maintain PC tumors or cancer stem cells in the BM (Winter et al., 2012).

Many acute infections lead to depletion of peripheral Treg cells (Benson et al., 2012; Oldenhove et al., 2009), and this

reduction in Treg populations in the BM correlates with the loss of pre-existing long-lived PCs. While evidence is presented that supports the idea that Treg cells maintain PCs, the underlying mechanism is unclear. The close association of Treg cells and PCs suggests there may be communication between these populations through cell contact or via soluble factors. Alternatively, the extended contacts between Treg cells and CD11c-YFP<sup>+</sup> cells in the BM may be relevant to PC maintenance. Thus, the close association of BM PCs with CD11c-YFP<sup>+</sup> cells is consistent with the idea that PC survival is dependent on constitutive signals through CD80/CD86 (Rozanski et al., 2011). Although DCs are required for Treg homeostasis (Darrasse-Jèze et al., 2009; Hänig and Lutz, 2008; Salomon et al., 2000), Treg cells regulate the activation of DC (Liu et al., 2009; Veldhoen et al., 2006) and transcriptional profiling indicated that BM Treg cells express an effector phenotype and pathways that are relevant to the modulation of DC function (Burzyn et al., 2013a, 2013b; Feuerer et al., 2010). Indeed, when Treg cells did not express CTLA-4, there were increased BM PCs suggesting that this pathway can limit local PC activity. However, these data have to be interpreted with care as the increased plasmablasts and PCs in the periphery of Foxp3<sup>Cre</sup>/CTLA4<sup>fllox/fllox</sup> mice may contribute to the increased PC number in the BM (Matheu et al., 2015; Wing et al., 2008, 2014). In the absence of a specific means to deplete or inactivate BM Treg cells, it is difficult to exclude the possibility that systemic effects of Treg cell deficiencies impact on BM PC numbers. An alternative explanation may be that inflammation leads to a simultaneous, but non-causal loss of both PCs and Treg cells, while delivery of IL-2 complexes may promote the survival or proliferation of other cell types required for the support of these populations. Regardless, we favor the possibility that the ability of Treg cells to interact with DCs may have a dual role: allowing Treg cells to contribute to the maintenance of the PC population, but also to limit PC activities.

The alterations observed in the BM during acute infection have been characterized in response to numerous pathogens, (Glatman Zaretsky et al., 2014), and the work presented here highlights effects on pre-existing PC populations, that appears to be common to infection or vaccination (Ng et al., 2014; Slocombe et al., 2013; Xiang et al., 2007). While the alterations in BM PCs are transient, the benefit of this disruption is unclear. It has been suggested that because the PC niche is of limited size, this process may allow newly created PCs to displace pre-existing PCs in order to expand the repertoire of the BM PC population to ensure that recent systemic pathogens are represented (Radbruch et al., 2006; Xiang et al., 2007). The observation that PCs removed from the BM undergo rapid apoptosis indicates that PCs are not intrinsically long lived and require constant signals for maintenance; however, our studies do not distinguish whether disruption of the niche leads to PC death or evacuation as a consequence of competition with plasmablasts (Odendahl et al., 2005). While the mechanisms by which PCs contract and subsequently are restored in the BM are uncertain, they may include cell death, a selection mechanism for the repopulation of the niche, and/or expansion of memory B cell populations (Bernasconi et al., 2002; DiLillo et al., 2008; Ng et al., 2014).

The success of rituximab highlights the clinical benefits of B cell depletion for many disease states, although this approach does not affect PC populations (Edwards and Cambridge, 2006). There are antibody-mediated disorders (allergy, lupus, and rheumatoid arthritis) where eliminating pre-existing PCs could have value (Edwards and Cambridge, 2006; Radbruch et al., 2006). However, as PCs express few traditional B cell markers, downregulate most surface receptors, including the BCR and MHC class II, and are radio-resistant, there are few treatments to deplete established PCs (Clatworthy, 2011; Lu et al., 2011). The studies presented here offer a model to understand the factors that govern homeostasis and maintenance of PCs and suggest an unexpected role for Treg cells in the processes that support PC populations, in addition to perhaps limiting PC activity. Understanding the PC-Treg cell interaction could provide strategies to treat immune-mediated diseases in which long lived PCs are involved.

## EXPERIMENTAL PROCEDURES

### Animals

C57BL/6J mice were obtained from Taconic or Sage at 6–8 weeks of age. BAC transgenic BLIMP1-YFP mice were originally from E. Meffre (Yale University). CD11c-YFP mice were provided by Michel Nussenzweig (Rockefeller University). Foxp3-GFP mice were from Vijay Kuchroo (Harvard University). Foxp3-DTR mice were obtained from Alexander Rudensky (Memorial Sloan Kettering). BLIMP1-GFP mice were originally from Stephen Nutt (WEHI). Foxp3-IRES-Cre BALB/c CTLA4<sup>lox/lox</sup> mice were previously described (Wing et al., 2008). All mouse strains were housed, maintained, and bred under specific pathogen-free conditions at the University of Pennsylvania, Harvard University, Osaka University, or UCSD animal facilities. All experimental procedures with mice were approved by the Institutional Animal Care and Use Committee of the respective institution.

### Infections, Immunizations, and Irradiation

*T. gondii* tachyzoites of the Prugniaud strain were maintained *in vitro* by serial passage in human fibroblasts monolayers and parasites were isolated via serial needle passage and filtration through 5  $\mu$ m filters. Mice were infected intraperitoneally (i.p.) with  $10^4$  parasites in 100  $\mu$ L PBS. NP-ovalbumin (Biosearch Technologies) with a conjugation ratio of 16–18 was emulsified in alum and 50  $\mu$ g/mouse was injected i.p. To make IL-2 complexes, 1.5  $\mu$ g carrier-free rIL-2 (eBioscience) was incubated with 25  $\mu$ g anti-IL-2 (JES6-1A12, eBioscience) for 5 min at room temperature before injection. Control mice received 25  $\mu$ g Rat IgG (BioXcell). Foxp3-DTR mice were treated with DT on days 0, 1, 4, and 6 for Treg cell depletion. For mixed BM chimeras, WT or CD11c-YFP recipient mice were irradiated with 1,000 Rd. After 6 hr,  $2 \times 10^6$  cells isolated from BM of Foxp3-GFP and CD11c-YFP (1:1) or only Foxp3-GFP (100%) donor mice were transferred *iv* to recipient mice. Mice were provided sulfamethoxazole in the drinking water for 2 weeks after irradiation. After 8–12 weeks of reconstitution, recipient mice were imaged as described.

### Cell Purification and Flow Cytometry

Briefly, mouse spleens and BM were collected and dissociated through a 40- $\mu$ m strainer to produce a single cell suspension. Red blood cells were lysed using 0.86% ammonium chloride (Sigma). Cells were counted, washed in flow cytometry buffer (1% BSA [Sigma], 2 mM EDTA [Invitrogen] in PBS). Fc receptors (FcR) were blocked with 2.4G2 (BioXCell) and Rat IgG (Invitrogen) while staining with McAb at 4° for 30 min. Intracellular staining was carried out using the eBioscience kit. Samples were acquired using a FACSCanto II or LSRFortessa flow cytometer (BD Biosciences) and analyzed with FlowJo software (Tree Star). The following monoclonal antibody (mAb) against mouse antigens were used as fluorescein isothiocyanat (FITC), PE, PerCP-Cy5.5, PE-Cy7, allophycocyanin (APC), APC-ef780, Pacific blue, AF450, AF700, PE Texas Red, or biotin conjugates: Fc $\epsilon$ RI, AA4.1, CD23, CD43, IgM, Fc $\gamma$ RII/III, ckit, Sca-1, CD34, CD138,

CD45R (B220; RA3-6B2), CD278 (ICOS; C398.4A), IgD (11-26) (eBioscience), CD4 (RM4-5), CD8a (53–6.7), CD95 (Fas; Jo2), CXCR5 (2G8) (BD Biosciences), CD3 (17A2), CD19 (6D5), Foxp3, CTLA-4, Siglec F, CD11b, F4/80, TCR $\beta$ , Gr-1, Fc $\epsilon$ RI, and CD279 (PD-1; J43) (Biolegend). Biotinylated antibodies were detected using PerCP-Cy5.5- (BD Biosciences) or APC-conjugated streptavidin (eBioscience). FITC or biotin-conjugated PNA was obtained from Vector Laboratories. Plots shown are on a Logicle scale.

### Microarray Profiling and Analysis

C57BL/6<sup>Foxp3-IRES-gfp</sup> mice were bred in specific-pathogen-free facilities at Harvard Medical School. BM Treg cells (GFP<sup>+</sup>CD25<sup>hi</sup>CD4<sup>+</sup>TCR $\beta$ <sup>+</sup>CD19<sup>-</sup>CD8 $\alpha$ <sup>-</sup>TCR $\gamma$  $\delta$ <sup>-</sup>CD11b<sup>-</sup>CD11c<sup>-</sup>NK1.1<sup>-</sup>Gr-1<sup>-</sup>Ter-119<sup>-</sup>) were triple-sorted (necessary to eliminate contamination from dominant myeloid populations) into Trizol from pools of two to three mice. Splenic Tregs were double-sorted from the same mice. Profiling was performed on Affymetrix MoGene 1.0 ST microarray as described (Sitrin PMID 23650440). Data were analyzed with the Multiplot Studio and Gene-E modules from GenePattern. Principal component analysis-based 3D plotting was generated with the Population PCA module (<http://cbdm.hms.harvard.edu/LabMembersPages/SD.html>).

### Histology

Mouse femurs were isolated and placed in 10% buffered formalin (Sigma). Tissues were paraffin embedded, sectioned, and stained with H&E, and slides were reviewed by a board-certified veterinary anatomic pathologist. Identification of PCs was based on characteristic morphologic features, including eccentrically located round to oval nuclei with coarsely clumped chromatin peripheral to the nuclear envelope, moderate amounts of deep purple cytoplasm with a distinct perinuclear clearing, indicative of a well-developed Golgi apparatus. Immature (e.g., band) neutrophils were identified as round cells with amphiphilic cytoplasm and horseshoe-shaped nuclei. Mature neutrophils were identified as round cells with pale eosinophilic cytoplasm and nuclei containing hyper-condensed chromatin with multiple segmentations.

### Microscopy

Mice were anaesthetized and maintained at core temperature of 37°C. Thinned-skull and open-skull surgery were performed as described previously (Nimmerjahn et al., 2005; Yang et al., 2010). For preparation of the spleen for imaging, mice were euthanized by CO<sub>2</sub> asphyxiation, the spleen was removed and immediately transferred into an imaging chamber (Warner Instruments). Tissue was perfused with cRPMI (RPMI plus 10% fetal-calf serum, 1% pen/strep, 1% glutamine, 1% nonessential amino acids, and 0.1%  $\beta$ -mercaptoethanol), bubbled with 95% oxygen and 5% carbon dioxide, and maintained at 37°C. Imaging was performed using a Leica SP5 2-photon microscope system (Leica Microsystems) equipped with a picosecond or femtosecond laser (Coherent). GFP, YFP, RFP, and tdTomato were excited using laser light of 900 nm or as indicated. Images were obtained using a 20 $\times$  water-dipping lens. Fluorescence emission was separated by high efficiency custom dichroic mirrors and collected with four external non-descanned detectors. The use of all four detectors allowed the separation of GFP and YFP expressing cells and from autofluorescent cells, which appear in “orange” due to their detection in several channels simultaneously. The resulting images were analyzed with Volocity software (PerkinElmer). In these images, the frequency of interaction and the duration of interaction between cells were assessed manually using three-dimensional reconstructions of the acquired movies (Movies S4 and S8). Three to four mice from each of the BLIMP1-YFP, Foxp3-GFP BLIMP1-YFP dual reporter, Foxp3-GFP CD11c-YFP dual reporter, and BLIMP1-GFP CD11c-YFP dual reporter strains were imaged and analyzed at each of the indicated time points. Within the Foxp3-GFP BLIMP1-YFP dual reporter mice, 53 BM and 104 splenic Foxp3-GFP<sup>+</sup> cells were imaged and analyzed. Eighty-three BLIMP1-GFP cells were imaged and quantified in BLIMP1-GFP CD11c-YFP dual reporter mice. Within the Foxp3-GFP CD11c-YFP dual reporter mice, 68 Foxp3-GFP<sup>+</sup> cells were imaged and evaluated in the BM and 94 splenic Foxp3-GFP<sup>+</sup> cells were imaged and analyzed. Fifty-three Foxp3-GFP<sup>+</sup> cells in the BM and 150 splenic Foxp3-GFP<sup>+</sup> cells were analyzed in BM chimeras that were made as described in the text. For transfer of total splenic T cells, splenocytes were isolated from naive mice and enriched for CD8<sup>+</sup> T cells by negative selection (Miltenyi). These cells were then activated and

expanded in vitro for 3 days with plate bound  $\alpha$ CD3(1  $\mu$ g/mL)/ $\alpha$ CD28(1  $\mu$ g/mL) and IL-2 (200 U/mL<sup>-1</sup>) before transfer of 1 million cells to recipient Foxp3-GFP mice.

For clearing and imaging bones were fixed in 4% PFA for 6 hr and immersed in 30% sucrose O/N before cryopreservation in OCT (Fisher Scientific) as previously described (Acar et al., 2015). Bones were bisected longitudinally on a cryostat, washed in PBS and blocked overnight in staining solution (10% DMSO, 0.5% IgePal630, and 5% donkey serum) plus FcR block. Tissues were incubated with labeled antibodies against GFP (Life Technologies), CD150 (Clone TC15-12F12.2, Biolegend), and Laminin 1+2 (polyclonal rabbit anti-mouse, Abcam) in staining solution for 3 days and then washed with PBS for 1 day. For clearing, half bones were dehydrated by incubation in increasing ethanol concentrations and overnight incubation in BABB solution (1:2, benzyl alcohol:benzyl benzoate). Images were obtained using a Zeiss LSM710 confocal microscope and analyzed using Velocity 6.3 (Perkin Elmer) software. These experiments were performed two to ten times on a total of greater than ten mice per image and representative images were selected.

### ELISA

Levels of NP-specific serum IgG1 were determined by ELISA. Immulon 4HBX plates (Thermo Fisher Scientific) were coated overnight at 4°C with 5  $\mu$ g NP-BSA/mL (Biosearch Technologies), in 50  $\mu$ m/well. Plates were then blocked with 10% FBS and incubated with serial dilutions of sera, followed by a peroxidase-coupled anti-IgG1 conjugate (Southern Biotech) and SureBlue TMB or ABTS substrate (KPL).

### Statistical Analysis

Statistical significance was assessed using a Student's t test. A p value of  $p \leq 0.05$  was considered significant. Statistical analysis was performed using Prism software (GraphPad Software).

### ACCESSION NUMBERS

The accession number for the data reported in this paper is GEO: GSE76264.

### SUPPLEMENTAL INFORMATION

Supplemental Information includes Supplemental Experimental Procedures, three figures, one table, and ten movies and can be found with this article online at <http://dx.doi.org/10.1016/j.celrep.2017.01.067>.

### AUTHOR CONTRIBUTIONS

A.G.Z., C.K., F.D., J.B.W., R.G., D.G.A., J.S.S., S.C., A.I.W., and W.J.Q. performed experiments and analysis. J.B.E. provided pathology expertise. D.C.B. offered statistical analysis. D.B., J.E., D.A., M.P.C., S.S., L.L., and C.O.B. provided technical and intellectual guidance and commented on the manuscript. A.G.Z. and C.A.H. wrote the manuscript.

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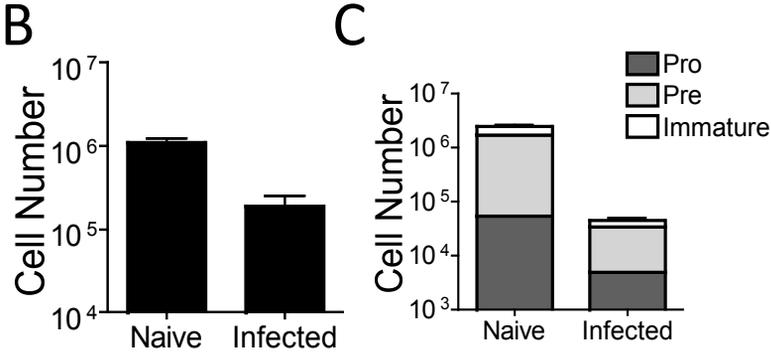
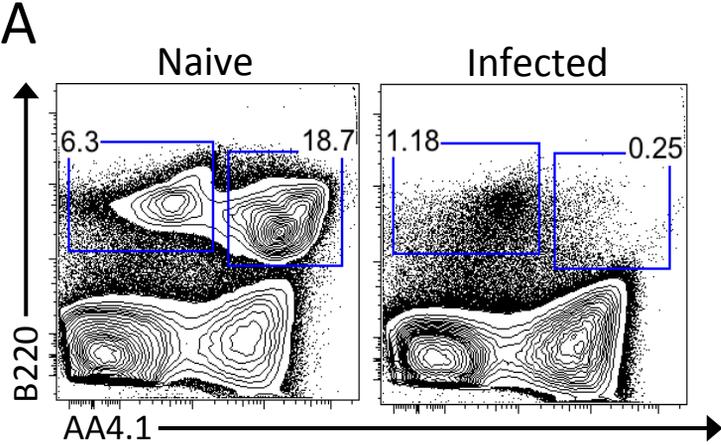
## **Supplemental Information**

### **T Regulatory Cells Support**

#### **Plasma Cell Populations in the Bone Marrow**

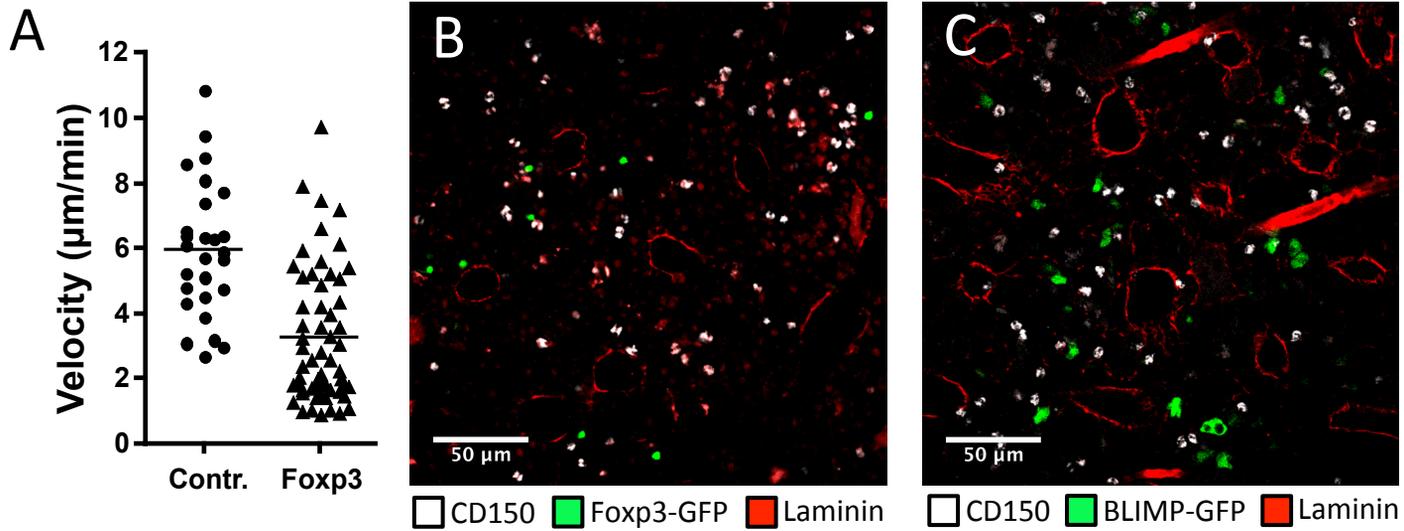
**Arielle Glatman Zaretsky, Christoph Konradt, Fabien Dépis, James B. Wing, Radhika Goenka, Daniela Gomez Atria, Jonathan S. Silver, Sunglim Cho, Amaya I. Wolf, William J. Quinn, Julie B. Engiles, Dorothy C. Brown, Daniel Beiting, Jan Erikson, David Allman, Michael P. Cancro, Shimon Sakaguchi, Li-Fan Lu, Christophe O. Benoist, and Christopher A. Hunter**

# Supplemental Figure 1



**Supplemental Figure 1. Related to Figure 1.**  
Developing B cell populations are reduced in the BM during infection. (A-C) BM from naïve or infected WT mice was evaluated by flow cytometry for the presence of pro, pre, immature (A and C), and mature recirculating (A-B) B cell populations.

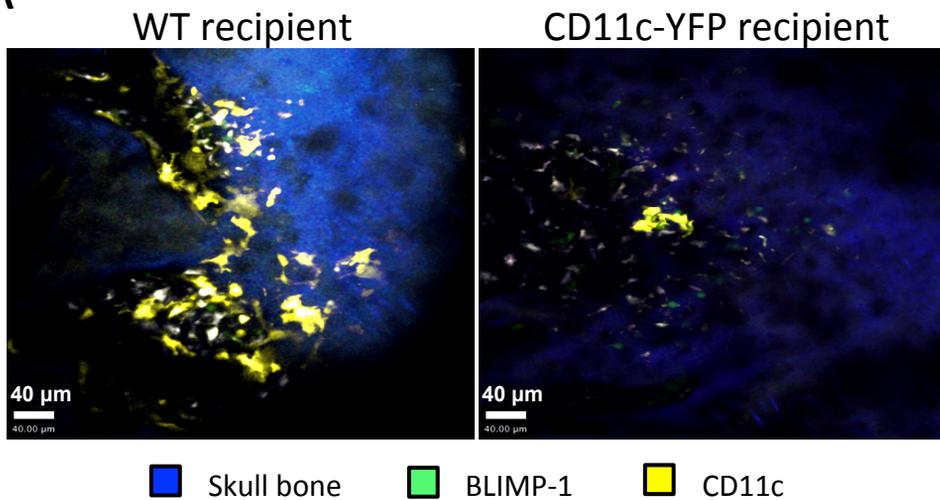
# Supplemental figure 2



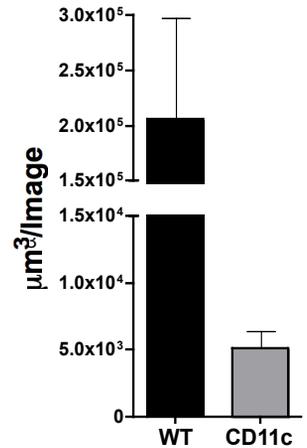
**Supplemental Figure 2. Related to Figure 2.** The cellular niches in the BM occupy an overlapping special region. **(A)** Splenic T cells were isolated from naïve mice. Cells were activated and expanded *in vitro* and transferred to Foxp3-GFP reporter mice. These mice were then imaged using intravital 2-photon microscopy of the skull BM. 3 mice were imaged in each of 2 experiments, with 2-3 fields containing transferred cells per mouse, for a total of 27 transferred cells recorded. **(B-C)** Femurs from Foxp3-GFP **(B)** and BLIMP-GFP **(C)** reporter mice were cleared, stained for laminin (red) and CD150 (white), and imaged by confocal microscopy.

# Supplemental figure 3

## A



## B



**Supplemental Figure 3. Related to Figure 3. (A and B)** CD11c-YFP<sup>+</sup> cells are primarily of hematopoietic origin. Recipient CD11c-YFP reporter or WT control mice were irradiated. WT recipients received CD11c-YFP and BLIMP1-GFP donor BM, thus CD11c-YFP cells were of hematopoietic origin. CD11c-YFP recipients received BLIMP1-GFP BM, making CD11c-YFP cells the radiation-resistant non-hematopoietic host cells. The BM of these recipient mice was imaged intravitaly to identify the origin of the CD11c-YFP<sup>+</sup> cells interacting with PC in the BM. There were an average of 15 cells per image in WT recipients and 2 per image in the reporter recipients, thus representative images are shown (A). The volume of YFP<sup>+</sup> cells in each of 8 randomly selected images per mouse was quantified (B).

Figure 4 Table S1

ProbeSetID	GeneSymbol	FC: [BM/SP]	P-value: BM.vs.SP
10389207	Ccl5	3.86	5.3E-04
10519983	Fgl2	3.73	5.3E-03
10345791	Il1rl1	3.44	4.9E-04
10590628	Ccr3	2.81	1.9E-03
10547590	Klrg1	2.54	6.5E-06
10531415	Cxcl10	2.40	1.4E-03
10503098	Lyn	2.33	2.6E-03
10486875	Frmd5	2.31	4.2E-04
10420308	Gzmb	2.28	1.0E-02
10590631	Ccr2	2.26	6.7E-03
10606058	Cxcr3	2.22	2.4E-04
10345807	Il18r1	2.22	7.0E-03
10531737	Hpse	2.21	3.3E-02
10493820	S100a6	2.10	5.5E-03
10494001	Tdpoz4	2.09	3.3E-02
10454198	Rnf125	2.06	3.3E-04
10487987	Lrrn4	2.05	3.7E-03
10443980	Myo1f	2.03	3.4E-03
10558580	Utf1	2.00	2.3E-03
10526832	LOC100504914	1.94	6.2E-04
10384974	Il9r	1.92	8.6E-03
10478633	Mmp9	1.91	3.9E-03
10493812	S100a4	1.89	3.4E-02
10368970	Prdm1	1.85	6.1E-03
10435704	Cd80	1.85	7.0E-05
10597575	Plcd1	1.82	1.3E-02
10454015	Ttc39c	1.82	1.1E-03
10544660	Osbpl3	1.82	7.1E-04
10371220	Gna15	1.81	2.7E-03
10450501	Tnf	0.55	1.4E-03
10406852	Cnn3	0.54	5.2E-03
10571344	D8Ertd82e	0.54	1.1E-02
10403413	Idi1	0.53	3.7E-03
10358408	Rgs1	0.53	3.2E-02
10583286	Gpr83	0.49	9.4E-04
10420362	Gjb2	0.49	2.0E-03
10363735	Egr2	0.49	2.3E-04
10454782	Egr1	0.47	1.9E-03
10450508	Lta	0.47	1.8E-03
10368199	Myb	0.43	1.9E-04
10472235	Dapl1	0.43	4.7E-03
10403825	Tcr-g-C	0.39	2.5E-03
10533182	Dtx1	0.33	1.3E-04
10381096	Igfbp4	0.30	3.2E-04
10407940	Tcr-g-V2	0.29	1.6E-04
10403821	Tcr-g-V3	0.28	6.0E-04

**Table S1. Related to Figure 4.** BM Treg cells exhibit a more effector-like phenotype than splenic Treg cells, with a higher expression of suppressor genes. Treg cells sorted from spleen and BM of naïve Foxp3-GFP<sup>+</sup> mice were evaluated by microarray. Cells were assessed for differentially regulated genes, listed here as fold change (FC) with p-value.

## Movies S1-S10

**Movie S1. Related to Figure 1. (A)** 3D projection of Plasma cells found in the endosteal region of the BM. Representative 360 degree rotational view of the BM using 2-Photon microscopy in a BLIMP1-YFP reporter mouse showing plasma cells (yellow), vasculature, stained with quantum dots (red) and the bone (blue).

At least 3 mice were imaged and a representative rotation is shown. **(B)** Intravital imaging showing that plasma cells are stationary in the BM of naïve mice.

Representative 20 min time lapse movie of PC (yellow) in the BM captured in a BLIMP1-YFP reporter mouse. Vasculature was stained with quantum dots (red).

At least 3 mice were imaged and a representative movie is shown.

**Movie S2. Related to Figure 2.** Foxp3<sup>+</sup> T regulatory cells are motile in the BM of naïve mice. Representative 20 min time lapse movie of Treg cells (green) in the BM captured in a Foxp3-GFP reporter mouse. Vasculature was stained with quantum dots (red). At least 3 mice were imaged.

**Movie S3. Related to Figure 2. (A)** Plasma cells and Treg cells are closely associated in the BM of naïve mice. Representative 20 min time lapse movie of PC (white) and Treg (green) cells in the BM captured in a BLIMP1-YFP/Foxp3-GFP dual reporter mouse. Vasculature was stained with quantum dots (red). 3 mice were imaged for these experiments, with 53 Foxp3-GFP cells recorded in the BM. **(B)** Plasma cells and Treg cells are closely associated in the BM of naïve mice. 3D projection of (A) for close up view of PC-Treg cell interactions.

Representative 20 min time lapse movie of PC (white) and Treg (green) cells in the BM captured in a BLIMP1-YFP/Foxp3-GFP dual reporter mouse. Vasculature was stained with quantum dots (red).

**Movie S4. Related to Figure 2.** Plasma cells and Treg cells are interacting in the BM of naïve mice. 360 degree rotation of a close up of Movie S3. PC (white) and Treg (green) cells in the BM captured in a BLIMP1-YFP/Foxp3-GFP dual reporter mouse. Vasculature was stained with quantum dots (red)

**Movie S5. Related to Figure 2.** Foxp3<sup>+</sup> T regulatory cells are highly motile with shorter interaction times in the spleens of naïve mice. Representative 20 min time lapse movie of Treg cells (green) and PC (white) in the spleen of a BLIMP1-YFP/Foxp3-GFP reporter mouse. 3 mice were imaged for these experiments, with 104 Foxp3-GFP cells recorded in the spleen.

**Movie S6. Related to Figure 2.** Activated CD8<sup>+</sup> T cells (Red) exhibit a higher velocity compared to Treg cells (green) in the BM. Representative 20 min time lapse movie of transferred CD8 T cells (red) and host Treg (green) cells in the BM captured in a Foxp3-GFP reporter mouse. 3 mice were imaged in each of 2 experiments, with 2-3 fields containing transferred cells per mouse, for a total of 27 transferred cells recorded.

**Movie S7. Related to Figure 2.** Plasma cells are associated with stromal cells in the BM. 360 degree rotational view of the BM using 2-Photon microscopy in a BLIMP1-GFP/CXCL12-RFP dual reporter mouse showing plasma cells (green) and stromal cells (red). 3 mice were imaged in these experiments, for a total of 44 BLIMP1-GFP+ cells.

**Movie S8. Related to Figure 3. (A)** Plasma cells and CD11c-YFP<sup>+</sup> cells are closely associated in the BM of naïve mice. Movie shows a z-stack in 2 µm sections through the BM of a BLIMP1-GFP/CD11c-YFP dual reporter mouse. Showing PC (green) interacting with CD11c<sup>+</sup> cells (yellow). Bone represented in blue. 4 mice were imaged, for a total of 83 BLIMP-GFP cells counted for quantification. **(B)** Plasma cells and CD11c-YFP<sup>+</sup> cells are closely associated in the BM of naïve mice. A 3D 360 degree rotational movie of the interaction between a PC (green) and a CD11c-YFP<sup>+</sup> (yellow) cell captured in the BM of a BLIMP1-GFP/CD11c-YFP dual reporter mouse.

**Movie S9. Related to Figure 3. (A)** Foxp3<sup>+</sup> T regulatory cells are highly motile and have brief interaction times with CD11c-YFP<sup>+</sup> cells in the spleens of naïve mice. Representative 20 min 3D time lapse movie of Treg cells in the spleens of a CD11c-YFP/Foxp3-GFP reporter mouse. 3 mice were imaged, for a total of 94 splenic Foxp3-GFP cells. **(B)** 360 degree rotational close up view of (A) showing that most Foxp3<sup>+</sup>T regulatory cells do not interact with CD11c-YFP<sup>+</sup> cells in the spleen

**Movie S10. Related to Figure 3. (A)** Foxp3<sup>+</sup> T regulatory cells are less mobile and have prolonged interaction times with CD11c-YFP<sup>+</sup> cells in the BM of naïve mice. Representative 20 min time lapse movie of Treg cells (green) in the BM of a CD11c-YFP/Foxp3-GFP reporter mouse. 3 mice were imaged, for a total of 68 BM Foxp3-GFP cells. **(B)** Foxp3<sup>+</sup> T regulatory cells are less mobile and have prolonged interaction times with CD11c-YFP<sup>+</sup> cells in the BM of naïve mice. Representative 3D projection of a 20 min time lapse movie of Treg cells (green) in the BM of a CD11c-YFP/Foxp3-GFP reporter mouse.